

INVESTICE DO ROZVOJE VZDELAVANI

How to analyze nucleic acid?

Název:

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Datum: 30.5.2014

Reg.č.projektu: CZ.1.07/2.3.00/20.0148

Název projektu: Mezinárodní spolupráce v oblasti "in vivo" zobrazovacích technik



How to analyze nucleic acid?

Sequence determination

- Blotting
- PCR
- Electrophoresis
- Microarray
- Sequencing

Structure determination

- NMR
- X-ray spectroscopy
- Circular dichroism

Quantification

- UV/VIS spectroscopy
- Electrochemical methods

Structure of nucleic acid



Structural motifs of nucleic acid

single strands	hairpins
duplex	stem-loops
A-Form helix	bulges
B-Form helix	Mismatches
Z-Form helix	DNA-RNA hybrid
parallel stranded	and complexes with:
triplexes	proteins and peptides
quadruplexes	ligands (drugs)
junctions	covalent
aptamers	intercalated
pseudoknots	groove binding

Nuclear magnetic resonance (NMR)

NMR is a highly developed and powerful spectroscopic technique that is valuable in the investigation of the structural, thermodynamic and kinetic properties of nucleic acids. The technique can be used to study DNA duplexes, triplexes, quadruplexes, hairpin loops, RNA duplexes and other secondary and tertiary RNA structures. Primarily, ¹H NMR is used in aqueous buffers with water suppression.

NMR should always be used in conjunction with other physical techniques such as circular dichroism (indicative of conformation, A, B, Z), Ultraviolet melting of duplexes, triplexes and quadruplexes (thermodynamic information), X-ray diffraction (high resolution structure analysis), and fluorescence resonance energy transfer (distance measurements).



Nuclei in a magnetic field absorb and re-emit electromagnetic radiation. This energy is at a specific resonance frequency which depends on the strength of the magnetic field and the magnetic properties of the isotope of the atoms; A key feature of NMR is that the resonance frequency of a particular substance is directly proportional to the strength of the applied magnetic field.

Typical NMR chemical shifts of base and sugar protons

NMR

Orientation of magnetic moments of nuclei







počet chemicky ekvivalentních jader v signálu

intenzita signálů

2D NMR



Structure of Nucleic Acid

2D homonuclear correlation spectroscopy



Very weak $J_{H1'-H2'}$ and strong $J_{H3'-H4'}$ cross-peaks correspond to *pure N-type conformation* (preferred conformation in RNA). Strong $J_{H1'-H2'}$ and weak $J_{H3' H4'}$ cross-peaks correspond to *pure S-type conformation*. $J_{H2'-H3_{,}}$ is similar in both states. Intermediate intensities indicate an equilibrium between N and S states.





Distance information (H8 - H1') determines the glycosidic torsion angle

The base can exist in 2 distinct orientations about the N-glycosidic bond. These conformations are identified as, **syn** and **anti**.

The anti conformation predominates.



Furthermore, although H8 resonances from nucleotides in the *anti* glycosidic conformation are in close contact to several sugar proton resonances, in the *syn* conformation no close contact to sugar protons is expected (except to the anomeric resonances).

X-ray crystallography

Advances in solid-phase oligonucleotide synthesis and purification have allowed a large number of oligonucleotides to be crystallized and their molecular structures solved. This has provided important information on both the B and A forms of DNA, confirming predictions made from earlier fibre-diffraction studies, and providing high-resolution information on the sequence dependence of nucleic acid structure.



crystal

reciprocal lattice

electron density map

model

When a crystal is placed in the path of an X-ray source it will generate a diffraction pattern, from which, with the help of some complicated mathematics, a three-dimensional picture of the unit cell and the molecules present can be generated.

X-ray crystallography

X-ray crystallography requires highly ordered crystals, and obtaining suitable crystals of nucleic acids is normally the slowest part of the process.

A highly concentrated solution of an oligonucleotide is required to provide a thermodynamic driving force for crystal formation. The solubility limit is exceeded by gradually removing water from the oligonucleotide solution using an external dehydrating agent, or diffusing a precipitant such as isopropanol into the solution. High-resolution structures of A, B and Z-DNA have been obtained by X-ray crystallography and the technique has been used to study DNA-drug complexes in atomic detail.

In most cases the resolution of the structure does not extend beyond 2 Å (0.2 nm), but this is sufficient to provide a reasonably clear picture of the heterocyclic bases and sugar-phosphate backbone.







Molecules with helical symmetry

- differential absorption of left and right circularly polarized light
- it is exhibited in the absorption bands of optically active chiral molecules
- when circularly polarized light passes through an absorbing optically active medium, the speeds between right and left polarizations differ ($c_L \neq c_R$) as well as their wavelength ($\lambda_L \neq \lambda_R$) and the extent to which they are absorbed ($\varepsilon_L \neq \varepsilon_R$). *Circular dichroism* is the difference $\Delta \varepsilon \equiv \varepsilon_L \varepsilon_R$.



Chromophores of NA



Structure of DNA









Thank you for your attention!



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