

# Fluorescence imaging for specific analysis of cancer cells

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Cancer is a serious disease that causes 25% of deaths in the developed countries. Significant impact on the cancer patients survival has early detection of this disease, therefore great attention is paid to its imaging. Fluorescence imaging represents powerful imaging method for the cell detection. For the successful detection of tumour cells, the specific targeting of fluorescence probes to the tumour tissue has a key role. Interesting materials enabling the imaging of tumour cells are fluorescence nanoparticles. For the accurate imaging, the NPs should be conjugated with targeting ligands and/or constructed as off-on probes.

**Keywords:** cancer cell; fluorescence; imaging; quantum dots

## 1. Introduction

Cancer is life-threatening disease, which causes nearly 7 million deaths every year worldwide and represents around 1 trillion dollars economic loss [1,2]. Cancer presents 25% of death caused in the developed countries [3]. But due to an early diagnosis and effective treatment, the mortality caused by this disease decreases and survival time increases [4]. The risk of dying from cancer decreased by 20% between 1991 and 2010 [3]. The cancer is diagnosed in the every third woman and every second man in the United States [5]. Cancer mortality in Czech Republic is about 20% [6,7]. Death rates continue to decline for all 4 major cancer sites (lung, colorectal, breast, and prostate), with lung cancer accounting for almost 40% of the total decline in men and breast cancer accounting for 34% of the total decline in women [4]. The most common causes of death are cancers of the lung, followed by colorectal, breast and stomach [7]. Early detection of cancer can significantly impact survival of cancer patients, so the regular screening is highly recommended [8-10].

## 2. Fluorescence imaging of cells

Optical methods are relatively cheap non-ionizing techniques based on the specific optical properties and are due to their variability, flexibility, specificity and sensitivity an important tool for non-invasive and objective diagnosis with still improving resolution [11-14]. Fluorescence imaging is the optical method based on the usage of fluorophores, the compounds, that can emit light after absorption of the appropriate wavelength light [15]. Fluorescence detection of cancer cells has the potential to be used in early cancer diagnosis [16]. The fluorescence of the dyes for imaging should be in the „tissue optical window“ spectral ranges between 650 and 900 nm [17] or in the infrared spectrum [18]. The long wavelength (far-red to NIR) fluorescent probes are advantageous for *in vivo* bioimaging because of minimum photo-damage to biological samples, deep tissue penetration, and minimum interference from background autofluorescence by biomolecules in the living systems. Therefore great attention is paid in the development of new long wavelength fluorescent probes [19].

### 3. Fluorescent probes

As smart molecular probes, organic compounds such as fluorescein or rhodamin are widely studied [20,21] and as an universal, genetically encoded fluorescent label, green fluorescent protein, is applied [22]. Promising imaging tools represent inorganic compounds such as nanoparticles (NPs) [23,24]. The NPs accumulate in tumour cells by themselves, or they have to be targeted to the cancer cell surface molecules [25]. Interesting NPs with very good fluorescence properties are QDs [26-28]. They have broad absorption spectrum and a narrow emission spectrum and are photostable [29,30]. Fluorescence properties of QDs can be successfully exploit for imaging of tumour cells as well as in situ investigations of tumour tissue [31,32]. Because of the content of heavy metals, the toxicity of QDs is discussed [33]. CdTe QDs conjugated with antibodies were successfully used for the cancer cells detection [34,35].

Technical developments in fluorescence imaging have enabled recent translation into investigational human studies [36]. NIR fluorescence imaging is in some applications already used in the clinic. Indocyanine green (ICG) is NIR fluorophore used primarily in angiography [37]. ICG enable noninvasive imaging of the lymphatic vasculature and discrimination of malignant from benign breast lesions [36]. ICG is useful for the hepatocellular carcinoma detection [38]. ICG encapsulated within poly-(allylamine) hydrochloride chains cross-linked ionically with sodium phosphate (ICG-NCs) and functionalized with anti-HER2 can be used as theranostic agent for optical imaging and also for the photodestruction of ovarian cancers invitro [39].

Tryptophan is investigated as the key native marker in cells to determine the level of metastasis competence in breast cell lines. The ratio of fluorescence intensity is associated with aggressiveness of the cancer cells [40]. The higher content of tryptophan was detected in the advanced metastatic cancer cell lines against the moderate metastatic and non aggressive cell lines [41].

Cancer cells can reduce of innocuous silver salts and spontaneously generate silver na-

noclusters (NCs), which has great fluorescence intensity. The formation of silver NCs also results in drastic reduction of tumour size and/or complete remission of the tumour [42]. Glutathione (GSH) can significantly and selectively enhance the fluorescence intensity of gold NCs. Gold NCs without GSH can selectively image the cancer cells. The liver cancer cells have much higher content of GSH, than other cell types. Therefore gold NCs enabled differentiation of cancer cells from normal ones [43].

For the targeting imaging, gold NCs conjugated with folic acid (FA) were utilized [44]. Folate receptor (FR) is overexpressed by a number of epithelial-derived tumors and minimally expressed in normal tissues. As FA is a high-affinity ligand to FR, and not produced endogenously, the FA-conjugated probes are specific for cancer cells imaging [45]. Folate-functionalized NPs are efficient imaging probes [46]. The overexpression of FR in 90–95% of epithelial ovarian cancers enabled the investigation of intraoperative tumor-specific fluorescence imaging in ovarian cancers surgery using an FR-targeted fluorescent probe [47]. Fluorescence imaging of cancer cells utilized in guidance surgery improves the tumour successful removal [48].

Low cytotoxic fluorescence probe comprises hydroxy-6-methyl-naphthalene-2-carbaldehyde was used for the detection of tyrosine kinase in cancer cells [49]. Lanthanum hexaboride  $\text{LaB}_6$  NPs coated with a carbon-doped silica ( $\text{C-SiO}_2$ ) shell to introduce a fluorescent property and improve stability and biocompatibility enabled fluorescent imaging and NIR-triggered photothermal therapy of cancer cells [50]. 2-[N-(7-nitrobenz-2-oxa-1,3-diazol-4-yl)amino]-2-deoxy-D-glucose (2-NBDG) was used as a tracer for detection of hypermetabolic circulating tumor cells (CTC) [51].

### 4. Turn off-on probes

For the sensitive and selective imaging without false positive results the „off-on“ fluorescence probes have been constructed. Polyethyleneimine-coated CdS/ZnS QDs (PEI-CdS/ZnS QDs) with the electrostaticly absorbed FA were turned off. In the presence of FR, the FA was desorbed and the fluorescence signal of QDs

was detected [45]. The fluorescence-quenching platform based on the biomineralized HAP (hydroxyapatite) has been also internalized for cancer cell detection [52]. Protein LAPTM4B is characteristic for a large number of cancer cells. Small peptide IHGHHIISVG (AP2H) is a targeting ligand for the LAPTM4B and therefore could be used for targeting the fluorescence probe to the cancer cells. Huang et al. constructed the turn-on probe consisted of the peptide and tetraphenylethylene (TPE), an aggregation-induced emission (AIE) fluorophore [21]. Another turn-on probe was designed via hydrogen-bond interaction between FA and carbon dots (CDs) with the passivating agent-poly(acrylate sodium) (PAAS). This probe could be used in the fluorescence-assisted surgical resection and real-time monitoring of the cells [16]. Acid phosphatase (ACP) is an important biomarker and indicator of prostate cancer. An „off-on“ probe for ACP detection consisted of a near-infrared mercaptopropionic acid (MPA)-capped CuInS<sub>2</sub> QDs [53].

## 5. Fluorescence *in situ* hybridization

Fluorescence *in situ* hybridization (FISH) with centromeric probes is used to detect chromosomal instability (CIN), which is observed in many cancers. The chromosome doubling could influence the tumours heterogeneity [54]. A multi-gene fluorescence *in situ* hybridization (M-FISH) was used to investigate gene copy number aberrations (CNAs) and it was found, that the gene copy number aberrations (CNAs) of cell cycle-regulated genes can be significant for prognosis in young breast cancer patients [55]. FISH is a useful technique for ALK gene rearrangement analysis and specification of the type of gene irregularities. ALK gene examination could be applied in histological and also in cytological samples. FISH was utilized in the FISH of samples from NSCLC patients [56].

## 6. Conclusions

Cancer is one of the most serious diseases worldwide. The most important for its successful treatment is early detection of tumour in the body. Methods for the fast and easy tumour detection are widely studied. Interesting appli-

cation represents fluorescence imaging. There are many possibilities of the fluorescence imaging of tumour cells. The most important for the use of fluorescence imaging in human medicine will be the necessity of the construction of the highly sensitive fluorescence detectors to detect the fluorophores deeply in the body.

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## Conflicts of Interest

The authors declare no conflict of interest.

## References

1. Fan, Z.; Senapati, D.; Singh, A.K.; Ray, P.C. Theranostic magnetic core-plasmonic shell star shape nanoparticle for the isolation of targeted rare tumor cells from whole blood, fluorescence imaging, and photothermal destruction of cancer. *Mol. Pharm.* 2013, 10, 857-866.
2. Sultana, S.; Khan, M.R.; Kumar, M.; Kumar, S.; Ali, M. Nanoparticles-mediated drug delivery approaches for cancer targeting: A review. *J. Drug Target.* 2013, 21, 107-125.
3. Siegel, R.; Ma, J.M.; Zou, Z.H.; Jemal, A. Cancer statistics, 2014. *CA-Cancer J. Clin.* 2014, 64, 9-29.
4. Siegel, R.; Naishadham, D.; Jemal, A. Cancer statistics, 2012. *CA-Cancer J. Clin.* 2012, 62, 10-29.
5. Siegel, R.; DeSantis, C.; Virgo, K.; Stein, K.; Mariotto, A.; Smith, T.; Cooper, D.; Gansler, T.; Lerro, C.; Fedewa, S., et al. Cancer treatment and survivorship statistics, 2012. *CA-Cancer J. Clin.* 2012, 62, 220-241.
6. Zaloudik, J. *Onkologicky vyzkum v ceske republice v souvislostech. Klinicka onkologie 2007*, 405-407.
7. Ferlay, J.; Steliarova-Foucher, E.; Lortet-Tieulent, J.; Rosso, S.; Coebergh, J.W.W.; Comber, H.; Forman, D.; Bray, F. Cancer incidence and mortality patterns in Europe: Estimates for 40 countries in 2012. *European Journal of Cancer* 2013, 49, 1374-1403.
8. Carter, H.B.; Albertsen, P.C.; Barry, M.J.; Etzioni, R.; Freedland, S.J.; Greene, K.L.; Holmberg, L.; Kantoff, P.; Konety, B.R.; Murad, M.H., et al. Early detection of prostate cancer: AUA guideline. *Journal of Urology* 2013, 190, 419-426.
9. Esserman, L.J.; Thompson, I.M.; Reid, B. Overdiagnosis and overtreatment in cancer an opportunity for improvement. *JAMA-J. Am. Med. Assoc.* 2013, 310, 797-798.
10. Horilova, J.; Cunderlikova, B.; Marcek Chorvatova, A. Time- and spectrally resolved characteristics of flavin fluorescence in u87mg cancer cells in culture. *Journal of biomedical optics* 2015, 20, 51017.
11. Blazkova, I.; Vaculovicova, M.; Eckschlager, T.; Stiborova, M.; Trnkova, L.; Adam, V.; Kizek, R.

- Study of fluorescence of doxorubicin in muscle tissue using highly sensitive fluorescence sensing. *Chem. Sensors* 2014, 4, 1-6.
12. Balas, C. Review of biomedical optical imaging-a powerful, non-invasive, non-ionizing technology for improving in vivo diagnosis. *Meas. Sci. Technol.* 2009, 20, 1-12.
  13. Stemmer, N.; Mehnert, J.; Steinbrink, J.; Wunder, A. Noninvasive fluorescence imaging in animal models of stroke. *Curr. Med. Chem.* 2012, 19, 4786-4793.
  14. Wouters, F.S.; Vermeer, P.J.; Bastiaens, P.I.H. Imaging biochemistry inside cells. *Trends Cell Biol.* 2001, 11, 203-211.
  15. Frangioni, J.V. In vivo near-infrared fluorescence imaging. *Curr. Opin. Chem. Biol.* 2003, 7, 626-634.
  16. Liu, Q.L.; Xu, S.H.; Niu, C.X.; Li, M.F.; He, D.C.; Lu, Z.L.; Ma, L.; Na, N.; Huang, F.; Jiang, H., et al. Distinguish cancer cells based on targeting turn-on fluorescence imaging by folate functionalized green emitting carbon dots. *Biosens. Bioelectron.* 2015, 64, 119-125.
  17. Pu, Y.; Tang, R.; Xue, J.P.; Wang, W.B.; Xu, B.G.; Achilefu, S. Synthesis of dye conjugates to visualize the cancer cells using fluorescence microscopy. *Appl. Optics* 2014, 53, 2345-2351.
  18. Rosenblum, L.T.; Kosaka, N.; Mitsunaga, M.; Choyke, P.L.; Kobayashi, H. Optimizing quantitative in vivo fluorescence imaging with near-infrared quantum dots. *Contrast Media & Molecular Imaging* 2011, 6, 148-152.
  19. Yuan, L.; Lin, W.; Zheng, K.; He, L.; Huang, W. Far-red to near infrared analyte-responsive fluorescent probes based on organic fluorophore platforms for fluorescence imaging. *Chemical Society Reviews* 2013, 42, 622-661.
  20. Chatterjee, K.; Zhang, J.Q.; Honbo, N.; Karliner, J.S. Doxorubicin cardiomyopathy. *Cardiology* 2010, 115, 155-162.
  21. Huang, Y.Y.; Hu, F.; Zhao, R.; Zhang, G.X.; Yang, H.; Zhang, D.Q. Tetraphenylethylene conjugated with a specific peptide as a fluorescence turn-on bioprobe for the highly specific detection and tracing of tumor markers in live cancer cells. *Chem.-Eur. J.* 2014, 20, 158-164.
  22. Chudakov, D.M.; Matz, M.V.; Lukyanov, S.; Lukyanov, K.A. Fluorescent proteins and their applications in imaging living cells and tissues. *Physiological Reviews* 2010, 90, 1103-1163.
  23. Andreadou, I.; Sigala, F.; Iliodromitis, E.K.; Papaefthimiou, M.; Sigalas, C.; Aligiannis, N.; Savvari, P.; Gorgoulis, V.; Papalabros, E.; Kremastinos, D.T. Acute doxorubicin cardiotoxicity is successfully treated with the phytochemical oleuropein through suppression of oxidative and nitrosative stress. *Journal of Molecular and Cellular Cardiology* 2007, 42, 549-558.
  24. Minotti, G.; Menna, P.; Salvatorelli, E.; Cairo, G.; Gianni, L. Anthracyclines: Molecular advances and pharmacologic developments in antitumor activity and cardiotoxicity. *Pharmacol. Rev.* 2004, 56, 185-229.
  25. Key, J.; Leary, J.F. Nanoparticles for multimodal in vivo imaging in nanomedicine. *International Journal of Nanomedicine* 2014, 9, 711-726.
  26. Zhu, Y.; Hong, H.; Xu, Z.P.; Li, Z.; Cai, W. Quantum dot-based nanoprobes for in vivo targeted imaging. *Curr. Mol. Med.* 2013, 13, 1549-1567.
  27. Wang, Y.C.; Hu, R.; Lin, G.M.; Roy, I.; Yong, K.T. Functionalized quantum dots for biosensing and bioimaging and concerns on toxicity. *ACS Appl. Mater. Interfaces* 2013, 5, 2786-2799.
  28. Moullick, A.; Blazkova, I.; Milosavljevic, V.; Fohlerova, Z.; Hubalek, J.; Kopel, P.; Vaculovicova, M.; Adam, V.; Kizek, R. Application of cdte/zns quantum dots in in vitro imaging of chicken tissue and embryo. *Photochem. Photobiol.* 2015, 91, 417-423.
  29. Hong, L.; Wang, Z.; Yuan, L.; Tan, J.H.; Wang, L.X.; Qu, G.B.; Zhang, D.Q.; Lin, R.H.; Liu, S.J. Subcellular distribution of cdse quantum dots (qds) in breast cancer cells. *J. Nanosci. Nanotechnol.* 2012, 12, 365-367.
  30. Gallo, J.; Garcia, I.; Genicio, N.; Penades, S. Cdte-based qds: Preparation, cytotoxicity, and tumor cell death by targeting transferrin receptor. *Part. Part. Syst. Charact.* 2014, 31, 126-133.
  31. Hu, W.Q.; Fang, M.; Zhao, H.L.; Yan, S.G.; Yuan, J.P.; Peng, C.W.; Yang, G.F.; Li, Y.; Li, J.D. Tumor invasion unit in gastric cancer revealed by qds-based in situ molecular imaging and multispectral analysis. *Biomaterials* 2014, 35, 4125-4132.
  32. Moullick, A.; Blazkova, I.; Milosavljevic, V.; Fohlerova, Z.; Hubalek, J.; Kopel, P.; Vaculovicova, M.; Adam, V.; Kizek, R. Application of cdte/zns quantum dots in in vitro imaging of chicken tissue and embryo. *Photochemistry and Photobiology* 2014.
  33. Lin, G.M.; Ding, Z.C.; Hu, R.; Wang, X.M.; Chen, Q.; Zhu, X.M.; Liu, K.; Liang, J.H.; Lu, F.Q.; Lei, D.L., et al. Cytotoxicity and immune response of cdse/zns quantum dots towards a murine macrophage cell line. *RSC Adv.* 2014, 4, 5792-5797.
  34. Wang, Z.Y.; Zong, S.F.; Chen, H.; Wang, C.L.; Xu, S.H.; Cui, Y.P. Sers-fluorescence joint spectral encoded magnetic nanoprobes for multiplex cancer cell separation. *Adv. Healthc. Mater.* 2014, 3, 1889-1897.
  35. Chang, B.X.; Yang, X.J.; Wang, F.; Wang, Y.S.; Yang, R.; Zhang, N.; Wang, B.Q. Water soluble fluorescence quantum dot probe labeling liver cancer cells. *J. Mater. Sci.-Mater. Med.* 2013, 24, 2505-2508.
  36. Sevick-Muraca, E.M. Translation of near-infrared fluorescence imaging technologies: Emerging clinical applications. *Annual Review of Medicine*, Vol 63 2012, 63, 217-231.
  37. Mielke, D.; Malinova, V.; Rohde, V. Comparison of intraoperative microscopic and endoscopic icg angiography in aneurysm surgery. *Neurosurgery* 2014, 10, 418-425.
  38. Morita, Y.; Sakaguchi, T.; Unno, N.; Shibasaki, Y.; Suzuki, A.; Fukumoto, K.; Inaba, K.; Baba, S.; Takehara, Y.; Suzuki, S., et al. Detection of hepatocellular carcinomas with near-infrared fluorescence imaging using indocyanine green: Its usefulness and limitation. *International Journal of Clinical Oncology* 2013, 18, 232-241.
  39. Bahmani, B.; Guerrero, Y.; Vullev, V.; Singh, S.P.; Kundra, V.; Anvari, B. Icg-loaded polymeric nanocapsules functionalized with



- anti-her2 for targeted fluorescence imaging and photodestruction of ovarian cancer cells. Reporters, Markers, Dyes, Nanoparticles, and Molecular Probes for Biomedical Applications V 2013, 8596.
40. Zhang, L.; Pu, Y.; Xue, J.P.; Pratavieira, S.; Xu, B.G.; Achilefu, S.; Alfano, R.R. Tryptophan as the fingerprint for distinguishing aggressiveness among breast cancer cell lines using native fluorescence spectroscopy. *Journal of biomedical optics* 2014, 19.
  41. Pu, Y.; Xue, J.P.; Xu, B.G.; Wang, W.B.; Gu, Y.Q.; Tang, R.; Achilefu, S.; Ackerstaff, E.; Koutcher, J.A.; Alfano, R.R. Investigation of native fluorescence spectral difference among prostate cancer cell lines with different risk levels. *Optical Biopsy Xi* 2013, 8577.
  42. Gao, S.P.; Chen, D.H.; Li, Q.W.; Ye, J.; Jiang, H.; Amatore, C.; Wang, X.M. Near-infrared fluorescence imaging of cancer cells and tumors through specific biosynthesis of silver nanoclusters. *Sci Rep* 2014, 4.
  43. Zhang, X.D.; Wu, F.G.; Liu, P.D.; Gu, N.; Chen, Z. Enhanced fluorescence of gold nanoclusters composed of hauc14 and histidine by glutathione: Glutathione detection and selective cancer cell imaging. *Small* 2014, 10, 5170-5177.
  44. Ding, C.Q.; Tian, Y. Gold nanocluster-based fluorescence biosensor for targeted imaging in cancer cells and ratiometric determination of intracellular ph. *Biosens. Bioelectron.* 2015, 65, 183-190.
  45. Zhang, Y.; Liu, J.M.; Yan, X.P. Self-assembly of folate onto polyethyleneimine-coated cds/zns quantum dots for targeted turn-on fluorescence imaging of folate receptor overexpressed cancer cells. *Anal. Chem.* 2013, 85, 228-234.
  46. Maity, A.R.; Saha, A.; Roy, A.; Jana, N.R. Folic acid functionalized nanoprobe for fluorescence-, dark-field-, and dual-imaging-based selective detection of cancer cells and tissue. *ChemPlusChem* 2013, 78, 259-267.
  47. van Dam, G.M.; Themelis, G.; Crane, L.M.A.; Harlaar, N.J.; Pleijhuis, R.G.; Kelder, W.; Sarantopoulos, A.; de Jong, J.S.; Arts, H.J.G.; van der Zee, A.G.J., et al. Intraoperative tumor-specific fluorescence imaging in ovarian cancer by folate receptor-alpha targeting: First in-human results. *Nature Medicine* 2011, 17, 1315-U1202.
  48. Metildi, C.A.; Felsen, C.N.; Savariar, E.N.; Nguyen, Q.T.; Kaushal, S.; Hoffman, R.M.; Tsien, R.Y.; Bouvet, M. Ratiometric activatable cell-penetrating peptides label pancreatic cancer, enabling fluorescence-guided surgery, which reduces metastases and recurrence in orthotopic mouse models. *Ann. Surg. Oncol.* 2015, 22, 2082-2087.
  49. Ahn, K.H.; Kim, D.; Kim, S.; Kim, K.H. Fluorescence probe useful for detecting tyrosine kinase for imaging cancer cells or tissues, comprises 3-hydroxy-6-methyl-naphthalene-2-carbaldehyde. WO2014181960-A2; KR2014133730-A, WO2014181960-A2 13 Nov 2014 C12Q-001/48 201478.
  50. Lai, B.H.; Chen, D.H. Lab6 nanoparticles with carbon-doped silica coating for fluorescence imaging and near-ir photothermal therapy of cancer cells. *Acta Biomater.* 2013, 9, 7556-7563.
  51. Cai, H.W.; Peng, F.Y. 2-nbdg fluorescence imaging of hypermetabolic circulating tumor cells in mouse xenograft model of breast cancer. *J. Fluoresc.* 2013, 23, 213-220.
  52. Zhang, Y.; Liu, W.; Banks, C.E.; Liu, F.; Li, M.; Xia, F.; Yang, X.L. A fluorescence-quenching platform based on biomineralized hydroxyapatite from natural seashell and applied to cancer cell detection. *Sci Rep* 2014, 4.
  53. Lin, Z.H.; Liu, Z.P.; Zhang, H.; Su, X.G. Near-infrared fluorescence probe for the determination of acid phosphatase and imaging of prostate cancer cells. *Analyst* 2015, 140, 1629-1636.
  54. Ito, H.; Oga, A.; Ikemoto, K.; Furuya, T.; Maeda, N.; Yamamoto, S.; Kawachi, S.; Itoh, H.; Oka, M.; Sasaki, K. Analysis of centromere signal patterns in breast cancer cells with chromosomal instability using image cytometry combined with centromere fluorescence in situ hybridization. *Cytom. Part A* 2014, 85A, 809-816.
  55. Li, C.Y.; Bai, J.C.; Hao, X.M.; Zhang, S.; Hu, Y.H.; Zhang, X.B.; Yuan, W.P.; Hu, L.P.; Cheng, T.; Zetterberg, A., et al. Multi-gene fluorescence in situ hybridization to detect cell cycle gene copy number aberrations in young breast cancer patients. *Cell Cycle* 2014, 13, 1299-1305.
  56. Wojas-Krawczyk, K.; Krawczyk, P.A.; Ramlau, R.A.; Szumilo, J.; Kozielski, J.; Kalinka-Warzechas, E.; Bryl, M.; Knopik-Nabrowicz, A.; Szychalski, L.; Szczesna, A., et al. The analysis of alk gene rearrangement by fluorescence in situ hybridization in non-small cell lung cancer patients. *Wspolczesna Onkol.* 2013, 17, 484-492.



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