

Hemagglutinin structure, membrane fusion and virus entry

Petr Michalek¹, Ludmila Krejcová¹, Vojtech Adam^{1,2} and Rene Kizek^{1,2*}

¹ Department of Chemistry and Biochemistry, Faculty of Agronomy, Mendel University in Brno, Zemedelska 1, CZ-613 00 Brno, Czech Republic, European Union;

² Central European Institute of Technology, Brno University of Technology, Technicka 3058/10, CZ-616 00 Brno, Czech Republic, European Union;

* Author to whom correspondence should be addressed; E-Mail: kizek@sci.muni.cz;

Received: 13.2.2015 / Accepted: 9.3.2015 / Published: 1.4.2015

Hemagglutinin (HA) is an antigenic glycoprotein, which is placed on the surface of the influenza viruses. It is responsible for binding the virus to the host cell, that is being infected. The name „hemagglutinin“ comes from the ability of protein to cause erythrocytes to agglutinate („clump together“). The process is like this: Hemagglutinin (HA) binds to the monosaccharide sialic acid which is present on the surface of its target host cells. The cell membrane then engulfs the virus through endocytosis and followed by formation of endosome. The cell then attempts to begin digesting the contents of the endosome by acidifying its interior and transforming it into a lysosome. When the pH decrease to 6.0, the HA molecule becomes partially unfold, and release a hydrophobic portion of peptide chain that was previously hidden. This so-called „fusion peptide“ acts like a molecular grapple hook for lock on the endosomal membrane. The rest of the HA molecule refolds into a new structure and pulls the endosomal membrane right up next to the viral membrane, causing the two to fuse together. When it happened, the viral RNA genome enters into the cell's cytoplasm.

1. Introduction

Influenza A viruses belong to the order Orthomyxoviridae and are responsible for significant annual morbidity and mortality. They are classified serologically based on the antigenic properties of their surface glycoproteins: the hemagglutinin (HA) and the neuraminidase (NA) [1]. To date, 18 HA subtypes, caused by antigenic shift, have been determined [2, 3]. These subtypes can be divided into 6 clades and two groups and this variability makes it difficult to effectively aim any drug against this structure. Furthermore, also antigenic drift can strengthen the ability to escape the virus from effective blockage [4].

2. Structure and function of hemagglutinin

HA is a type I transmembrane glycoprotein with a signal sequence that is removed post-translationally, a membrane anchor domain near the C terminus, and a short cytoplasmic

tail [5]. Its size is about 13.5 nm and a molecular weight of about 76 kDa [6]. HA is a target molecule for neutralizing antibodies, and therefore is considered as the major surface antigen [7]. Primary function of HA is the initiation of infection, HA also involved in the host cell recognition and binding of the virus to host cell receptor, which is composed of sialic acid [6, 8]. The virus-receptor binding is followed by virus entry into the host cell and release of viral RNA from the virion, which allows subsequent replication. Nowadays, H1 (H1N1) and H3 (H3N2) are the most widespread HA influenza subtypes in human population. Also other subtypes, which are typically occurred in waterfowl, may cause human infection or deaths such as H5 [9, 10], H7 [11], and H9 [12].

HA monomers are synthesized and assembled noncovalently to trimer in the endoplasmic reticulum, where also glycosylation occurs, and transported through the Golgi complex to the cell surface of infected cells as an uncleaved,

fusion-incompetent precursor HAO, which is proteolytically cleaved into two smaller subunits (HA1 and HA2) [13-16]. Disulphide bridges, which linkage HA1 and HA2 subunits, which were formed in viral replication in the HAO folding process, are cleaved [17, 18]. HAO cleavage can take place either in the Golgi apparatus or extracellularly (using enzymes produced by cells of the respiratory system), the process HAO cleavage is essential for infectivity of the virus particles [19, 20]. After cleavage of HAO precursors, three HA structures form a trimer, which has a mushroom shape [18]. Mushroom consists of antiparallel beta-sheet region HA1 subunits and elongated membrane-proximal domain (stem region) dominated by intertwined and interconnecting α -helices (HA2) [21]. Large membrane-distal, globular HA1 subunit, or the receptor binding region enables the binding of the virus to glycan receptors on host cells, which are formed by the sialic acid bounded by galactose [18, 22, 23].

3. Membrane fusion and virus entry

Sialic acid, linked to complex glycans on either glycoproteins or glycolipids, is the receptor for influenza binding. No significant conformational change of HA appears during receptor binding and virion is just attached to the cell surface. Fusion-inducing conformational change is activated by binding of one or more of protons, as the pH in the endosome goes lower [24]. Enveloped viruses enter cells through fusion of their viral membrane with a host cell membrane. This fusion process is thermodynamically favorable but kinetically very slow [25].

Influenza infection is initiated by the viral HA binding to sialic acid receptors on the surface of the host cell. It is widely accepted that the human-adapted HA subtypes preferentially bind to the $\alpha(2,6)$ -sialic acid linkage, whereas the avian-adapted HA subtypes preferentially bind to the $\alpha(2,3)$ -sialic acid linkage [26, 27]. Membrane fusion between host cell and influenza virus is a thermodynamically favorable process, but a high kinetic barrier is crossed as the two bilayers approach each other [28]. When bound to

the specific sialic acid receptor on the target cell, the influenza virion is endocytosed in coated pits and vesicles, and delivered to endosomes [17]. This process is cell-type dependent and influenza virus can enter the host cell using either clathrin-dependent and clathrin-independent endocytosis or by macropinocytosis [29-31]. Proton pumps in the membranes of endocytic vesicles induce an accumulation in protons and therefore lowering of the pH between 5 and 6, which is essential for HA cleavage and causes the HA1 'head' to separate from the HA2 'stem' and enables a set of HA2 conformational transformations [13, 32]. This change causes the exposure of the N terminus of HA2, known as the fusion peptide and is required to promote fusion between the viral envelope and the target membrane and therefore is essential for virus infection [33-35].

4. Conclusions

The infection process of HA has been well documented throughout the years. But, the understanding of the influenza viral attack mechanism is still crucial for designing of new antiviral therapeutics such as protease and fusion inhibitors and cross-neutralizing antibodies that interfere with the fusion process.

Acknowledgments

The financial support by FLUMED IP A 4.4 is greatly acknowledged.

Conflicts of Interest

The authors declare they have no potential conflicts of interests concerning drugs, products, services or another research outputs in this study. The Editorial Board declares that the manuscript met the ICMJE „uniform requirements“ for biomedical papers.

References

1. Bradley, K.C., et al., Analysis of Influenza Virus Hemagglutinin Receptor Binding Mutants with Limited Receptor Recognition Properties and Conditional Replication Characteristics. *Journal of Virology*, 2011. 85(23): p. 12387-12398.
2. Tong, S., et al., New World Bats Harbor Diverse Influenza A Viruses. *Plos Pathogens*, 2013. 9(10).
3. Wu, Y., et al., Bat-derived influenza-like viruses H17N10 and H18N11. *Trends in Microbiology*,

2014. 22(4): p. 183-191.
4. Yang, J., et al., Influenza A Virus Entry Inhibitors Targeting the Hemagglutinin. *Viruses-Basel*, 2013. 5(1): p. 352-373.
 5. Steinhauer, D.A., Role of hemagglutinin cleavage for the pathogenicity of influenza virus. *Virology*, 1999. 258(1): p. 1-20.
 6. Cheng, X., et al., Surface glycoproteins of influenza A H3N2 virus modulate virus replication in the respiratory tract of ferrets. *Virology*, 2012. 432(1): p. 91-98.
 7. Ducatez, M.F., et al., Feasibility of reconstructed ancestral H5N1 influenza viruses for cross-clade protective vaccine development. *Proceedings of the National Academy of Sciences of the United States of America*, 2011. 108(1): p. 349-354.
 8. Edinger, T.O., M.O. Pohl, and S. Stertz, Entry of influenza A virus: host factors and antiviral targets. *Journal of General Virology*, 2014. 95: p. 263-277.
 9. Nasreen, S., et al., Seroprevalence of Antibodies against Highly Pathogenic Avian Influenza A (H5N1) Virus among Poultry Workers in Bangladesh, 2009. *Plos One*, 2013. 8(9).
 10. Van Kerkhove, M.D., Brief literature review for the WHO global influenza research agenda - highly pathogenic avian influenza H5N1 risk in humans. *Influenza and Other Respiratory Viruses*, 2013. 7: p. 26-33.
 11. Guo, L., et al., Human Antibody Responses to Avian Influenza A(H7N9) Virus, 2013. *Emerging Infectious Diseases*, 2014. 20(2): p. 192-200.
 12. Bi, J.M., et al., Phylogenetic and Molecular Characterization of H9N2 Influenza Isolates from Chickens in Northern China from 2007-2009. *Plos One*, 2010. 5(9).
 13. Skehel, J.J. and D.C. Wiley, Receptor binding and membrane fusion in virus entry: The influenza hemagglutinin. *Annual Review of Biochemistry*, 2000. 69: p. 531-569.
 14. Stevens, J., et al., Structure and receptor specificity of the hemagglutinin from an H5N1 influenza virus. *Science*, 2006. 312(5772): p. 404-410.
 15. Boulay, F., et al., The influenza hemagglutinin precursor as an acid-sensitive probe of the biosynthetic-pathway. *Embo Journal*, 1987. 6(9): p. 2643-2650.
 16. DuBois, R.M., et al., The Receptor-Binding Domain of Influenza Virus Hemagglutinin Produced in *Escherichia coli* Folds into Its Native, Immunogenic Structure. *Journal of Virology*, 2011. 85(2): p. 865-872.
 17. Isin, B., P. Doruker, and I. Bahar, Functional motions of influenza virus hemagglutinin: A structure-based analytical approach. *Biophysical Journal*, 2002. 82(2): p. 569-581.
 18. Xu, R., et al., Structure, Receptor Binding, and Antigenicity of Influenza Virus Hemagglutinins from the 1957 H2N2 Pandemic. *Journal of Virology*, 2010. 84(4): p. 1715-1721.
 19. Garten, W. and H.D. Klenk, Understanding influenza virus pathogenicity. *Trends in Microbiology*, 1999. 7(3): p. 99-100.
 20. Leikina, E., et al., Reversible stages of the low-pH-triggered conformational change in influenza virus hemagglutinin. *Embo Journal*, 2002. 21(21): p. 5701-5710.
 21. Gamblin, S.J. and J.J. Skehel, Influenza Hemagglutinin and Neuraminidase Membrane Glycoproteins. *Journal of Biological Chemistry*, 2010. 285(37): p. 28403-28409.
 22. Rosenthal, P.B., et al., Structure of the haemagglutinin-esterase-fusion glycoprotein of influenza C virus. *Nature*, 1998. 396(6706): p. 92-96.
 23. Russell, R.J., et al., H1 and H7 influenza haemagglutinin structures extend a structural classification of haemagglutinin subtypes. *Virology*, 2004. 325(2): p. 287-296.
 24. Harrison, S.C., Mechanism of membrane fusion by viral envelope proteins, in *Virus Structure and Assembly*, P. Roy, Editor 2005, Elsevier Academic Press Inc: San Diego. p. 231-261.
 25. Chernomordik, L.V. and M.M. Kozlov, Protein-lipid interplay in fusion and fission of biological membranes. *Annual Review of Biochemistry*, 2003. 72: p. 175-207.
 26. Garcia-Sastre, A., Influenza Virus Receptor Specificity Disease and Transmission. *American Journal of Pathology*, 2010. 176(4): p. 1584-1585.
 27. Connor, R.J., et al., Receptor specificity in human, avian, and equine h2 and h3 influenza-virus isolates. *Virology*, 1994. 205(1): p. 17-23.
 28. Ivanovic, T., et al., Influenza-virus membrane fusion by cooperative fold-back of stochastically induced hemagglutinin intermediates. *Elife*, 2013. 2.
 29. Lakadamyali, M., M.J. Rust, and X.W. Zhuang, Ligands for clathrin-mediated endocytosis are differentially sorted into distinct populations of early endosomes. *Cell*, 2006. 124(5): p. 997-1009.
 30. de Vries, E., et al., Dissection of the Influenza A Virus Endocytic Routes Reveals Macropinocytosis as an Alternative Entry Pathway. *Plos Pathogens*, 2011. 7(3).
 31. Siczekarski, S.B. and G.R. Whittaker, Influenza virus can enter and infect cells in the absence of clathrin-mediated endocytosis. *Journal of Virology*, 2002. 76(20): p. 10455-10464.
 32. White, J.M. and I.A. Wilson, Anti-peptide antibodies detect steps in a protein conformational change - low-ph activation of the influenza-virus hemagglutinin. *Journal of Cell Biology*, 1987. 105(6): p. 2887-2896.
 33. Stegmann, T., J.M. White, and A. Helenius, Intermediates in influenza induced membrane-fusion. *Embo Journal*, 1990. 9(13): p. 4231-4241.
 34. Pak, C.C., M. Krumbiegel, and R. Blumenthal, Intermediates in influenza-virus pr/8 hemagglutinin-induced membrane-fusion. *Journal of General Virology*, 1994. 75: p. 395-399.
 35. Chen, J., J.J. Skehel, and D.C. Wiley, N- and C-terminal residues combine in the fusion-pH influenza hemagglutinin HA(2) subunit to form an N cap that terminates the triple-stranded coiled coil. *Proceedings of the National Academy of Sciences of the United States of America*, 1999. 96(16): p. 8967-8972.



The article is freely distributed under license Creative Commons (BY-NC-ND). But you must include the author and the document can not be modified and used for commercial purposes.